

Case Report

Regenerative Endodontic Management of an Immature Necrotic Premolar Using Advanced Platelet-Rich Fibrin

Sepideh Hosseini,¹ Nazanin Chitsaz ,² Mohammad Hassan Hamrah ,³ Donya Maleki,⁴ and Emad Taghizadeh ⁵

¹Department of Pediatric Dentistry, School of Dentistry, Guilan University of Medical Sciences, Rasht, Iran

²Department of Endodontics, School of Dentistry, Tehran University of Medical Sciences, Tehran, Iran

³Department of Public Health and Health Systems, Nagoya University Graduate School of Medicine, Nagoya, Japan

⁴Faculty of Dentistry, Guilan University of Medical Sciences, Rasht, Iran

⁵Department of Oral and Maxillofacial Surgery, School of Dentistry, Guilan University of Medical Sciences, Iran

Correspondence should be addressed to Emad Taghizadeh; emad_taghizadeh@gums.ac.ir

Received 22 September 2022; Revised 16 December 2022; Accepted 19 January 2023; Published 31 January 2023

Academic Editor: Hamdi Cem Gungor

Copyright © 2023 Sepideh Hosseini et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Regenerative endodontic management is a feasible treatment for immature teeth with periapical radiolucency and necrotic pulp that simplifies continued root creation. Among the most prevalent health problems in an immature root is dental pulp necrosis, which is caused by caries, improper endodontic treatments, and trauma. Necrosis of the dental pulp can affect long-term tooth survival and preservation and serve as a source of bacteria infecting the periapical tissue and the maxillofacial space. Here, we report on the application of advanced platelet-rich fibrin plus (A-PRF+) therapy, as a regenerative endodontic treatment (RET), in a 12-year-old with necrotic pulp and asymptomatic apical periodontitis. Over a 24-month follow-up post-treatment, we observed resolving of symptoms and a complete root formation with considerable periapical healing.

1. Introduction

Pulp necrosis in immature teeth leads to the incomplete development of roots, consisting of thin walls and open apices, making endodontic management challenging [1]. The pulp necrosis etiology in immature permanent teeth can include trauma, caries, and the existence of dental anomalies including dens evaginatus or dens invaginatus [2].

Immature teeth with apical periodontitis and pulp necrosis make challenges for endodontic management owing to the incomplete development of roots, thin root walls, and open apices [1]. For these cases, the conventional treatment modality includes apexification with single-step mineral trioxide aggregate (MTA) or the application of calcium hydroxide (CaOH) paste as an apical barrier to obtain apex closure. These two approaches have the drawback of restricting normal physiological root development, which

leads to thin and fragile roots [2]. A more recent therapeutic approach for such cases involves regenerative endodontic treatment (RET) in which root development is continued, conferring the advantages of vitality and maturation [3].

Several case studies have been published on immature necrotic teeth, indicating the potential of RET in stimulating the continuous formation of root length and width and preserving their structural integrity [4–6]. Using RET, healing of apical periodontitis is induced, restoring normal physiological functions of the pulp [7]. Continuous root development cannot be obtained with an apexification technique [8, 9].

More recently, studies have been performed on an autogenous material, platelet-rich fibrin (PRF), which serves as an osteoconductive scaffold with integrated growth factors for stimulating the cells for a regenerative response. While this technique has been utilized in dentistry for repairing different lesions and regeneration of dental and oral tissues, it can also

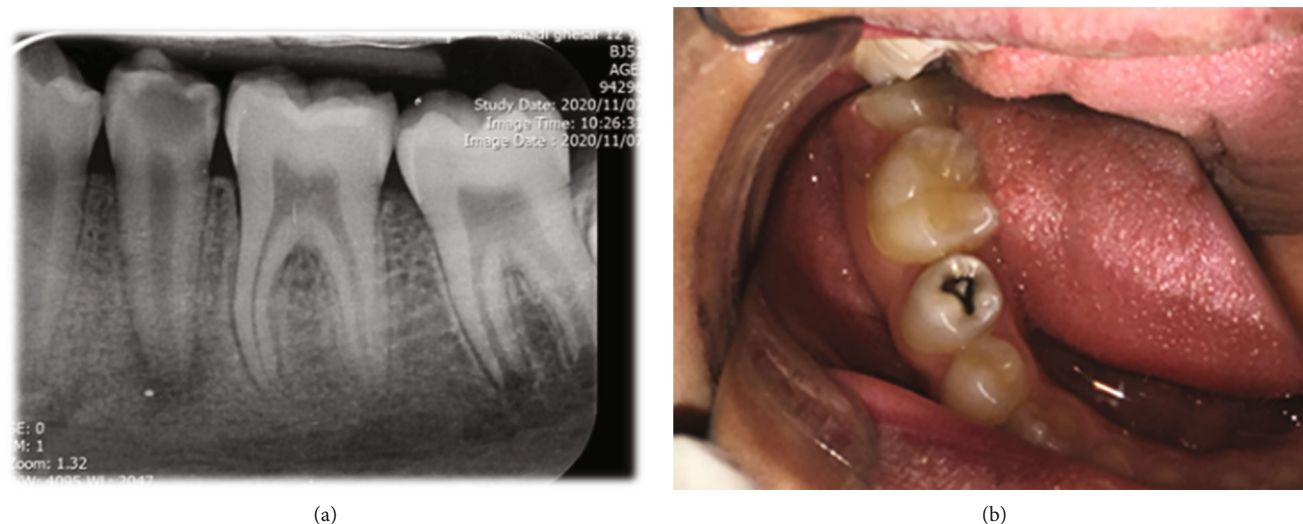


FIGURE 1: (a) The initial periapical radiograph showing prominent tubercles due to dens evaginatus, periapical radiolucency, and deep caries in tooth #20 with an immature apex; (b) preoperative photograph and caries in tooth #20; the intact tubercle can be observed on the occlusal surface of this tooth.

be employed as a scaffold for pulp regeneration in an immature necrotic tooth [10].

The present case report describes the management of a non-vital immature premolar tooth causing symptomatic apical periodontitis and necrosis of the pulp.

2. Case Presentation

The patient was an 11-year-old boy referred to the Department of Endodontic Dentistry of Tehran University of Medical Science, Tehran, Iran, with a complaint of spontaneous pain in the lower left premolar (tooth #20).

General dental, medical, and traumatic histories were obtained, and no systemic disease was noted. Extensive caries was revealed by clinical examination on the occlusal surface as well as sensitivity to palpation and percussion and dens evagination addressed as a differential diagnosis for this case. A negative response was obtained by the vitality test with electric pulp testing (EPT) (Digitest 3 Parkell, USA) and Endo-Frost cold spray (Roeko, Coltene Whaledent, Langenau, Germany), which represented no mobility. In the radiographic examination, an immature root was found along with deep caries, open apex, and periapical radiolucency. Additional caries was not found in the other teeth (Figure 1).

Necrotic pulp was diagnosed with symptomatic apical periodontitis. REP was proposed as the best feasible option for treatment. A full explanation was provided to the patient and his family about the risks and benefits of REP, and informed consent for the procedure was obtained.

An inferior alveolar nerve block was given with 2% lidocaine and 1:100,000 epinephrine (Daroupakhsh, Tehran, Iran) on the first visit. Caries removal and access cavity were conducted with diamond burs (Dentalree, USA) followed by rubber dam isolation. An apex locator (Woodpecker, Foshan, China) along with radiography (XGenus,

De Götzen SRL, Varese, Italy) was used to verify the working length. To irrigate the canals, 20 ml of 1.5% sodium hypochlorite (NaOCl) with ultrasonic passive activation was used. This was followed by irrigation with normal saline and drying by paper point (Dentplus, Choonchong, Korea). After applying CaOH into the canal, the access cavity was sealed temporarily with RMGI (Fuji IX, GC Corporation, Tokyo, Japan).

The second session was appointed after 3 weeks. The patient did not have any complications. Using 3% mepivacaine, the tooth was anesthetized without a vasoconstrictor (Daroupakhsh, Tehran, Iran). The temporary restorative material was removed after isolation with a rubber dam. Normal saline was used to wash Triple antibiotic paste (TAP) out of the canal. The canal was irrigated thoroughly with 20 ml of 17% Ethylenediamide tetraacetic acid (EDTA). PRF was prepared by drawing 10 ml of whole blood and centrifugation at 1300 rpm for 8 minutes. After drying the canals, a 40-K file was inserted 2 mm beyond the apex in the canal to initiate bleeding (Dentsply Maillefer, Ballaigues, Switzerland). The advanced platelet-rich fibrin plus (A-PRF+) was injected into the root canal to a level of 3 mm under the cemento-enamel junction (CEJ). To reduce the discoloration effect of MTA, a dentin bonding agent was placed on the dentinal walls. Ortho MTA (MTA-Ang; A Angelus, Londrina, PR, Brazil) was placed as a coronal barrier. A moist cotton pellet was placed over the MTA in the pulp chamber, and the access cavity was temporized again with GC1 filling (Figure 2).

A week later, during the third appointment, the glass ionomer and cotton pellet were removed. By setting MTA, restoration of the tooth was performed with composite (Clearfil AP-X, Kuraray Medical, Tokyo, Japan).

During follow-ups at months 3, 6, 12, and 24, the tooth remained asymptomatic with no sign of resorption, clinically or radiographically. Healing of the periapical lesion was

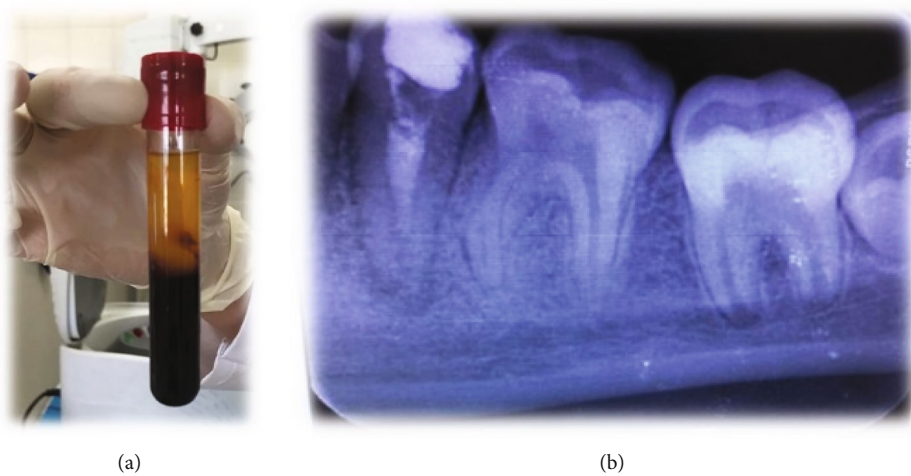


FIGURE 2: (a) Preparation of A-PRF+; (b) placing MTA on A-PRF+ and a temporary filling.

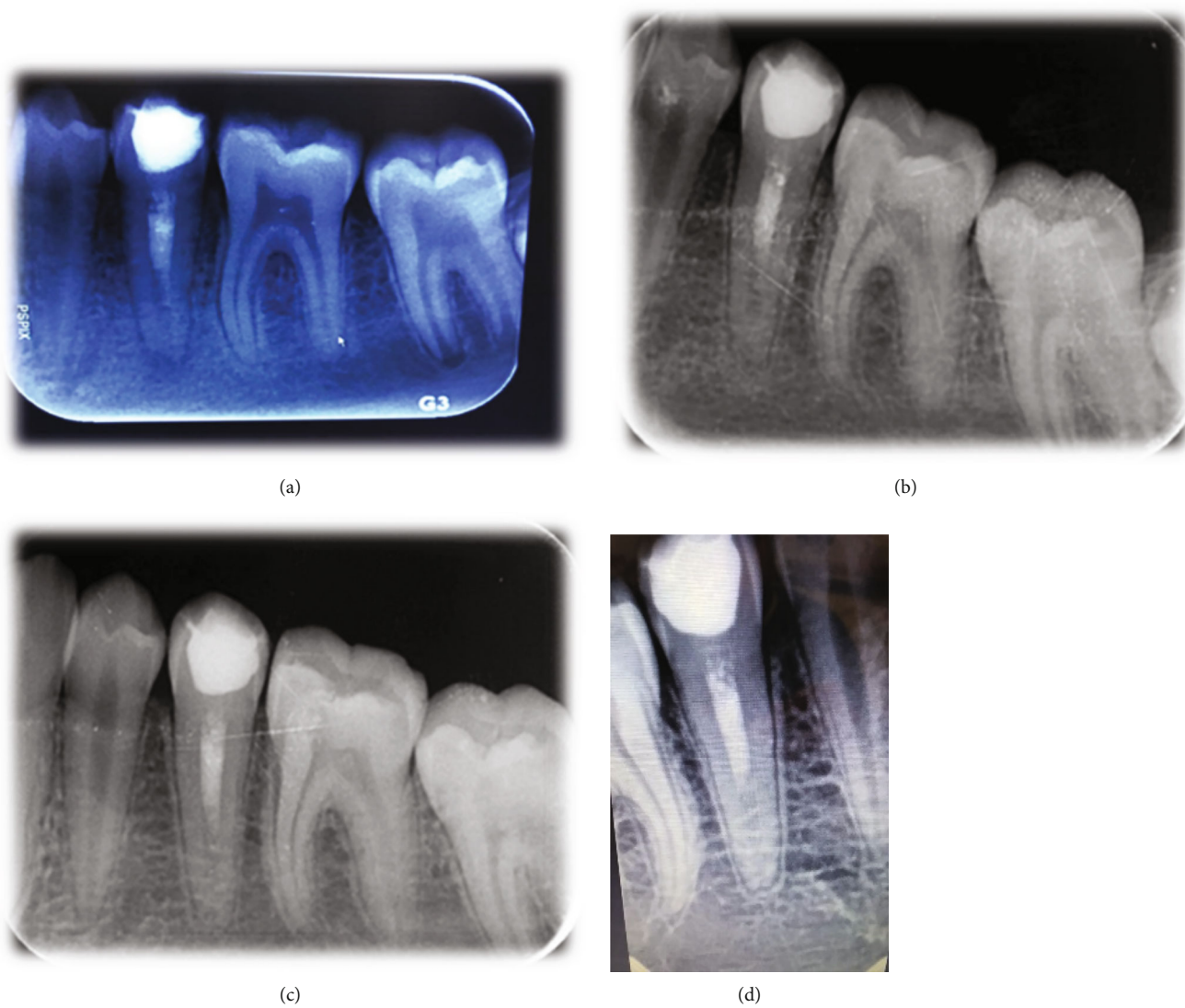


FIGURE 3: The periapical radiograph during follow-ups at months 3 (a), 6 (b), 12 (c), and 24 (d).

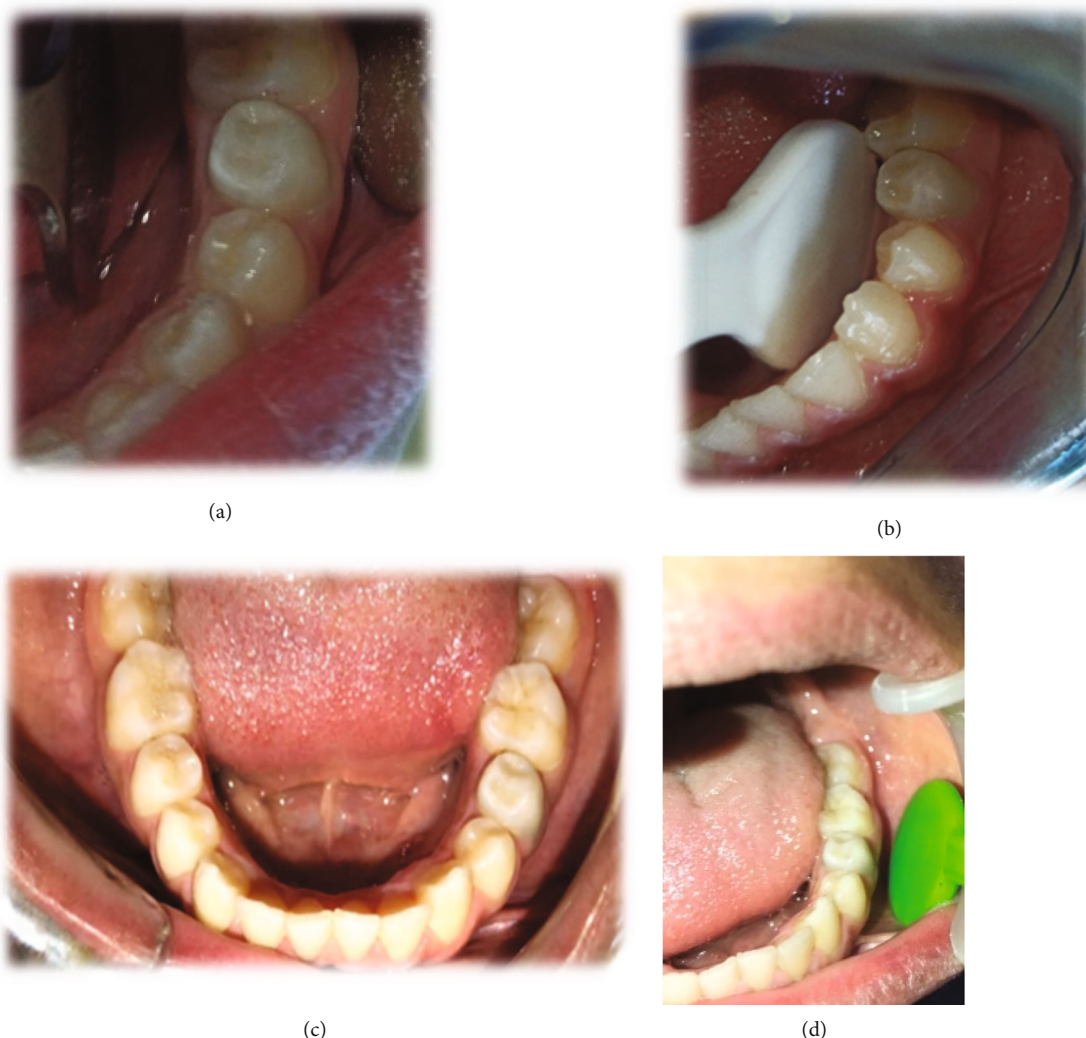


FIGURE 4: The postoperative photograph during follow-ups at months 3 (a), 6 (b), 12 (c), and 24 (d).

illustrated through a radiographic assessment, which showed incremented root thickness, length, and width, indicating the apical closure (Figures 3 and 4).

3. Discussion

This report described the administration of RET on immature pulp necrosis of a permanent premolar tooth with periapical radiolucency. The process conducted for this subject involved the placement of a freshly prepared PRF+ membrane inside the canal. From clinical experience, incremental placement of PRF fragments is more convenient compared to the placement of a membrane [11]. An MTA was directly placed over PRF to achieve a coronal seal [12, 13]. Satisfactory radiographic and clinical results were obtained over a 24-month follow-up.

Apexification and RET are two treatment modalities for an immature tooth with pulp necrosis. Apexification requires the intracanal application of CaOH, which should be replaced every 3 months [14, 15]. The long-term application of an intracanal medication and the development of

defects in the root walls owing to the CaOH's porous attributes increase the possibility of root fracture. On the other hand, RET, a shorter treatment, increases the root thickness and length and results in complete root development in a brief period [16–18]. MTA has been extensively used in the last decade for the apexification of non-vital immature teeth. Although it has several setbacks such as tooth discoloration and weakening of the dentin wall, it reduces the treatment time and is more effective than CaOH [19], and therefore, it can be considered as a treatment option to save the tooth. In this case, the American Association of Endodontists (AAE) recommendations were followed for irrigation protocols and the application of CaOH medication inside the root canal for 3 weeks [20].

Platelet products have great potential in regenerative medicine owing to their potency in releasing and storing biologically active materials, regulating the innate immune response, and combatting infection [21]. PRF has been presented as an autogenous source of blood growth factors, serving as a tool for tissue regeneration in modern medicine [22]. It was introduced in 2001 [23], and since then it has

been utilized extensively in dentistry for various procedures, with demonstrated effectiveness in treating gingival recessions, extraction socket management, intrabody defect regeneration, and sinus elevation [24–26]. One of its main advantages is the obtaining of immune-compatible growth factors at a low cost without the use of anti-coagulants [17]. Moreover, PRF has been associated with superior results in the regeneration of the pulp-dentin complex in endodontic treatments [10]. Reducing the centrifugation g-force is shown to increase the total leukocyte numbers within PRF matrix scaffolds, a modification known as A-PRF+ [27]. According to Fujioka's study, growth factors are released at significantly higher levels by A-PRF+ compared to L-PRF or A-PRF [22]. The decision on the use of A-PRF+ for RET in the present case was made based on the above information.

According to a meta-analysis, the mean success rate after the first year for apical closure or reduction has been 85% for both PRP (85.1%) and PRF (85.2%). The same value for root lengthening was 64.2% (PRP) and 74.1.2% (PRF). The periapical lesion healing response as well as dentinal wall thickening rates was 100% for both PRP and PRF [28]. However, Kobayashi et al. examined the release of various growth factors from PRP, PRF, and A-PRF+ for 15 minutes, 59 minutes, 8 hours, 1 day, 3 days, and 10 days and found that A-PRF+ released the maximum amount of growth factors over a longer duration when compared to PRF or PRP, which would be beneficial for regenerative procedures [29]. Moreover, other studies conducted on immature necrotic teeth have demonstrated the potential of RET in stimulating continuous root length and width formation and preserving their structural integrity [4–6]. The most important advantage of RET is, therefore, the continued root development, which cannot be obtained using an apexification technique [8, 9].

The tooth was deemed clinically asymptomatic, and there was a complete root formation with considerable periapical healing radiographically and a successful outcome. In addition to the limitation of case report in term of its essence, it should be mentioned that it was not possible to confirm the vitality of the pulp after regeneration due to the lack of access to laser Doppler flowmetry and pulse oximeter.

4. Conclusion

Regenerative endodontic treatment with A-PRF+ was effective in the treatment of apical periodontitis and immature permanent premolar with pulp necrosis. For 24 months, follow-up appointments were conducted, in which the tooth was deemed clinically asymptomatic with a complete root formation and considerable periapical healing, demonstrated radiographically.

Data Availability

Data supporting this research article are available from the corresponding author or first author on reasonable request.

Conflicts of Interest

The authors declare that they have no conflicts of interest.

References

- [1] P. E. Murray, F. Garcia-Godoy, and K. M. Hargreaves, "Regenerative endodontics: a review of current status and a call for action," *Journal of Endodontics*, vol. 33, no. 4, pp. 377–390, 2007.
- [2] J. Andreasen, B. Farik, and E. Munksgaard, "Long-term calcium hydroxide as a root canal dressing may increase risk of root fracture," *Dental Traumatology*, vol. 18, no. 3, pp. 134–137, 2002.
- [3] G. T. Huang, W. Sonoyama, Y. Liu, H. Liu, S. Wang, and S. Shi, "The hidden treasure in apical papilla: the potential role in pulp/dentin regeneration and bioroot engineering," *Journal of Endodontia*, vol. 34, no. 6, pp. 645–651, 2008.
- [4] M. Alasqah, S. I. R. Khan, K. Alfouzan, and A. Jamleh, "Regenerative endodontic management of an immature molar using calcium hydroxide and triple antibiotic paste: a two-year follow-up," *Case Reports in Dentistry*, vol. 2020, 2020.
- [5] J. Lin, Q. Zeng, X. Wei et al., "Regenerative endodontics versus apexification in immature permanent teeth with apical periodontitis: a prospective randomized controlled study," *Journal of Endodontia*, vol. 43, no. 11, pp. 1821–1827, 2017.
- [6] B. Thibodeau and M. Trope, "Pulp revascularization of a necrotic infected immature permanent tooth: case report and review of the literature," *Pediatric Dentistry*, vol. 29, no. 1, pp. 47–50, 2007.
- [7] American Association of Endodontists, *Glossary of Endodontic Terms*, p. 47, American Association of Endodontists, Chicago, IL, USA, 8th edition, 2012.
- [8] S. Koç and M. Del Fabbro, "Does the etiology of pulp necrosis affect regenerative endodontic treatment outcomes? A systematic review and meta-analyses," *The Journal of Evidence-Based Dental Practice*, vol. 20, no. 1, p. 101400, 2020.
- [9] T. Jeeruphan, J. Jantararat, K. Yanpiset, L. Suwannapan, P. Khewsawai, and K. M. Hargreaves, "Mahidol study 1: comparison of radiographic and survival outcomes of immature teeth treated with either regenerative endodontic or apexification methods—a retrospective study," *Journal of Endodontics*, vol. 38, no. 10, pp. 1330–1336, 2012.
- [10] E. Borie, D. G. Oliví, I. A. Orsi et al., "Platelet-rich fibrin application in dentistry: a literature review," *International Journal of Clinical and Experimental Medicine*, vol. 8, no. 5, pp. 7922–7929, 2015.
- [11] D. Keswani and R. K. Pandey, "Revascularization of an immature tooth with a necrotic pulp using platelet-rich fibrin: a case report," *International Endodontic Journal*, vol. 46, no. 11, pp. 1096–1104, 2013.
- [12] M. Torabinejad and M. Turman, "Revitalization of tooth with necrotic pulp and open apex by using platelet-rich plasma: a case report," *Journal of Endodontia*, vol. 37, no. 2, pp. 265–268, 2011.
- [13] E. J. Fischer, D. E. Arens, and C. H. Miller, "Bacterial leakage of mineral trioxide aggregate as compared with zinc-free amalgam, intermediate restorative material, and super-EBA as a root-end filling material," *Journal of Endodontia*, vol. 24, no. 3, pp. 176–179, 1998.

- [14] M. After, "Apexification: a review," *Dental Traumatology*, vol. 21, no. 1, pp. 1–8, 2005.
- [15] G. J. Huang, "Apexification: the beginning of its end," *International Endodontic Journal*, vol. 42, no. 10, pp. 855–866, 2009.
- [16] F. Banchs and M. Trope, "Revascularization of immature permanent teeth with apical periodontitis: new treatment protocol?," *Journal of Endodontia*, vol. 30, no. 4, pp. 196–200, 2004.
- [17] N. Shah, A. Logani, U. Bhaskar, and V. Aggarwal, "Efficacy of revascularization to induce apexification/apexogenesis in infected, nonvital, immature teeth: a pilot clinical study," *Journal of Endodontia*, vol. 34, no. 8, pp. 919–925, 2008.
- [18] L. Goyal, "Clinical effectiveness of combining platelet rich fibrin with alloplastic bone substitute for the management of combined endodontic periodontal lesion," *Restorative Dentistry and Endodontics*, vol. 39, no. 1, pp. 51–55, 2014.
- [19] S. N. M. P. Sockalingam, M. S. A. A. Awang Talip, and A. S. I. Zakaria, "Maturogenesis of an immature dens evaginatus non-vital premolar with an apically placed bioceramic material (EndoSequence Root Repair Material®): an unexpected finding," *Case Reports in Dentistry*, vol. 7, no. 2018, 2018.
- [20] American Association of Endodontics, "American Association of Endodontics clinical considerations for a regenerative procedure," May 2018, <https://www.aae.org/specialty/wpcontent/uploads/sites/2/2017/06/currentregenerativeendodonticconsiderations.pdf>.
- [21] V. Pavlovic, M. Ciric, V. Jovanovic, and P. Stojanovic, "Platelet rich plasma: a short overview of certain bioactive components," *Open Medicine*, vol. 11, no. 1, pp. 242–247, 2016.
- [22] M. Fujioka-Kobayashi, R. J. Miron, M. Hernandez, U. Kandam, Y. Zhang, and J. Choukroun, "Optimized platelet-rich fibrin with the low-speed concept: growth factor release, biocompatibility, and cellular response," *Journal of Periodontology*, vol. 88, no. 1, pp. 112–121, 2017.
- [23] J. Choukroun, F. Adda, C. Schoeffler, and A. Vervelle, "Opportunities in implant dentistry: PRF (French)," *Implantodontie*, vol. 42, p. e62, 2001.
- [24] D. R. Hoaglin and G. K. Lines, "Prevention of localized osteitis in mandibular third-molar sites using platelet-rich fibrin," *International Journal of Dentistry*, vol. 2013, 2013.
- [25] S. Aroca, T. Keglevich, B. Barbieri, I. Gera, and D. Etienne, "Clinical evaluation of a modified coronally advanced flap alone or in combination with a platelet-rich fibrin membrane for the treatment of adjacent multiple gingival recessions: a 6-month study," *Journal of Periodontology*, vol. 80, no. 2, pp. 244–252, 2009.
- [26] R. Padma, A. Shilpa, P. A. Kumar, M. Nagasri, C. Kumar, and A. Sreedhar, "A split mouth randomized controlled study to evaluate the adjunctive effect of platelet-rich fibrin to coronally advanced flap in Miller's class-I and II recession defects," *Journal of Indian Society of Periodontology*, vol. 17, no. 5, pp. 631–636, 2013.
- [27] S. Ghanaati, P. Booms, A. Orlowska et al., "Advanced platelet-rich fibrin: a new concept for cell-based tissue engineering by means of inflammatory cells," *The Journal of Oral Implantology*, vol. 40, no. 6, pp. 679–689, 2014.
- [28] P. E. Murray, "Platelet-rich plasma and platelet-rich fibrin can induce apical closure more frequently than blood-clot revascularization for the regeneration of immature permanent teeth: a meta-analysis of clinical efficacy," *Frontiers in Bioengineering and Biotechnology*, vol. 6, no. 6, p. 139, 2018.
- [29] E. Kobayashi, L. Flückiger, M. Fujioka-Kobayashi et al., "Comparative release of growth factors from PRP, PRF, and advanced-PRF," *Clinical Oral Investigations*, vol. 20, no. 9, pp. 2353–2360, 2016.